



HaDEA Service Contract 20197409

Provision of EU networking and support for public health reference laboratory functions for antimicrobial resistance in *Salmonella* species and *Campylobacter* species in human samples







Webinar

Guidance document on internal quality control schemes for reference antimicrobial susceptibility testing and detection of genetic determinants of antimicrobial resistance for *Salmonella* and *Campylobacter* isolates from human samples

Wednesday, 22 March 2023, 14:00-15:00 CET

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Virtual Housekeeping

Please **turn off your cameras and microphones** unless you're speaking – this will help with bandwidth and maximise audibility.

Do frequently **use the chat function** to share your views, comments and challenges. Keep the chat constructive, respectful and on topic!

If you wish to make a comment for e.g. the discussion, please use the 'Raise hand' function.









Meeting agenda

- 1. Presentation of the guidance document
- Background and aim
- IQC in general
- IQC for AST

2. Discussion







BACKGROUND AND AIM









- Collect the most recently available information from different regulatory agencies and other sources
- Aid the NRLs in optimizing their current methods and implementing molecular methods
- Provide concrete examples of techniques for Internal Quality Control
- Allow the NRLs to easily provide guidance or training to local national laboratories
- Describe the standardized / recommended methods for antimicrobial susceptibility testing

Reliable and accurate results for diagnostics and surveillance purposes of Salmonella and Campylobacter

Data that are comparable within Europe for surveillance purposes





BACKGROUND AND AIM

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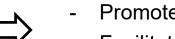




ISO standards

o ISO 15189

o ISO/IEC 17025



- Promote internal quality and competence
- Facilitate internal and external comparibility of results

EQA and accreditation

- External quality assessment exercises
- Strategy for accreditation



 Increase confidence regarding accuracy of results







ISO STANDARDS – ISO 15189

ISO standards

o **ISO 15189**

o ISO/IEC 17025

EQA and accreditation

- o External quality assessment exercises
- \circ Strategy for accreditation









ISO STANDARDS – ISO 15189

ISO 15189:2022

"Medical laboratories - Requirements for quality and competence"

Deep re-organization of the document compared with :2012 version

| :2012 | :2022 | |
|-------|-------|--|
| 1. | 1. | |
| 2. | 2. | |
| 3. | 3. | |
| 4. | 4. | |
| 5. | 5. | |
| | 6. | |
| | 7. | |
| | 8. | |
| | | |







ISO STANDARDS – ISO 15189







ISO STANDARDS – ISO 15189







ISO STANDARDS – ISO 15189

Example: 6) Resource requirements; 6.5) Equipment calibration and metrological traceability









ISO STANDARDS – ISO 15189

Example: 7) Process requirements; 7.3) Examination processes







ISO STANDARDS – ISO 15189

Example: 8) Management system requirements; 8.4) Control of records







ISO STANDARDS – ISO/IEC 17025

ISO standards



o ISO/IEC 17025

EQA and accreditation

- External quality assessment exercises
- Strategy for accreditation







ISO STANDARDS – ISO/IEC 17025

ISO/IEC 17025:2017

"General requirements for the competence of testing and calibration laboratories"







ISO STANDARDS – ISO/IEC 17025







ISO STANDARDS – ISO/IEC 17025

Now more similar to ISO 15189:2022







EXTERNAL QUALITY ASSESSMENT EXERCISES





EQA and accreditation

- External quality assessment exercises
- Strategy for accreditation







EXTERNAL QUALITY ASSESSMENT EXERCISES

Examples:

EARS-Net EQA / ECDC's EQA on AST / UK NEQAS / ESfEQA / Labquality / Oneworld Accuracy

Recommended within the ISO standards







STRATEGY FOR ACCREDITATION

ISO standards



EQA and accreditation

- External quality assessment exercises
- Strategy for accreditation









STRATEGY FOR ACCREDITATION

International Laboratory Accreditation Cooperation (ILAC)

^L Specifically designated body - national institution

^L Evaluation and accreditation of reference/local laboratories













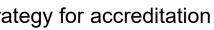


EQA and accreditation

External quality assessment exercises Ο



Strategy for accreditation 0











European guidance on AST methods:

European Committee on Antimicrobial Susceptibility Testing (EUCAST)

Recommendations:

- Broth microdilution or disk diffusion for AST
- Other methods (agar dilution / gradient strips) are not recommended due to lack of harmonisation and high variability
- Regularly confirming warnings and new breakpoint tables







Phenotypic antimicrobial susceptibility testing

- Broth microdilution
- \circ Disk diffusion
- \circ Detection of β -lactamases

Molecular detection of antimicrobial resistance

- PCR protocols
- Whole-genome sequencing







PHENOTYPIC AST – BROTH MICRODILUTION

Phenotypic antimicrobial susceptibility testing

- Broth microdilution
- \circ Disk diffusion
- \circ Detection of β -lactamases

Molecular detection of antimicrobial resistance

- o PCR protocols
- Whole-genome sequencing







PHENOTYPIC AST – BROTH MICRODILUTION

Standard protocol – ISO 20776-1:2019

 "Susceptibility testing of infectious agents and evaluation of performance of antimicrobial susceptibility test devices -Part 1: Broth micro-dilution reference method for testing the in vitro activity of antimicrobial agents against rapidly growing aerobic bacteria involved in infectious diseases"

EUCAST documents

- Clinical breakpoint tables
- Warnings page
- Visual guides (e.g. how to determine MIC endpoints)







PHENOTYPIC AST – BROTH MICRODILUTION

ISO 20776-1:2019







PHENOTYPIC AST – BROTH MICRODILUTION

ISO 20776-1:2019

- How to prepare stock and working solutions of antimicrobial agents, the broth medium and the microdilution trays
- Two methods for obtaining the bacterial inoculum: the broth culture method and the direct colony suspension method
 - Final concentration of 5 x 10⁵ CFU/ml
- How to inoculate, incubate and read the minimum inhibitory concentrations (MIC) on the microdilution trays
- Lists of situations that require special attention, including the adjustment of medium composition or incubation conditions for certain bacterial species and for certain antimicrobials









PHENOTYPIC AST – BROTH MICRODILUTION

Examples of special situations (ISO 20776-1 + EUCAST)

- For Salmonella spp.
 - Do not add surfactants to the medium when testing colistin
 - Adjust the zinc concentration of the broth medium for testing of carbapenems

- For *Campylobacter* spp.
 - Use MH-F broth (MH broth supplemented with lysed horse blood and β -NAD)
 - Incubate under the special conditions of 41± 1°C during 24 hours in microaerobic environment, and extension of incubation time up to 40-48 hours might be necessary if growth is insufficient







PHENOTYPIC AST – BROTH MICRODILUTION

- Use of control strains
 - List from the Clinical Laboratory Standards Institute (CLSI) (available on the document CLSI M100 "Performance Standards for Antimicrobial Susceptibility Testing")
 - List from EUCAST (available on the document "Routine and extended internal quality control for MIC determination and disk diffusion as recommended by EUCAST")

Salmonella spp.

Always: Escherichia coli ATCC 25922

Colistin: mcr-1-positive E. coli NCTC 13846

β-lactams+ β-lactamase inhibitors: *E. coli* ATCC 35218, others

Campylobacter spp.

Always: Staphylococcus aureus ATCC 29213

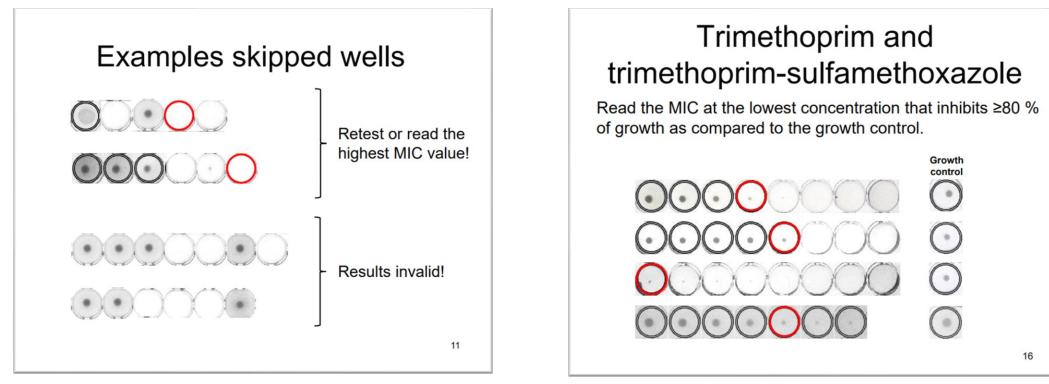






PHENOTYPIC AST – BROTH MICRODILUTION

EUCAST visual guidelines



EUCAST reading guide for broth microdilution, Version 4.0. Växjö, Sweden: 2022.



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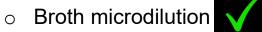






PHENOTYPIC AST – DISK DIFFUSION

Phenotypic antimicrobial susceptibility testing





- **Disk diffusion** Ο
- Detection of β -lactamases Ο

Molecular detection of antimicrobial resistance

- PCR protocols
- Whole-genome sequencing Ο







PHENOTYPIC AST – DISK DIFFUSION

Standard protocol – EUCAST protocol

o "Antimicrobial susceptibility testing - EUCAST disk diffusion method. Version 11.0, January 2023"

EUCAST documents

- o Clinical breakpoint tables
- Warnings page
- Visual guides (e.g. how to confirm adequate growth and determine zone diameters)







PHENOTYPIC AST – DISK DIFFUSION

EUCAST protocol

| Contents | | Page | |
|----------|---|------|--|
| | Changes from previous version | | |
| | Abbreviations and Terminology | | |
| 1 | Introduction | 5 | |
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| 3 | Preparation of inoculum | 8 | |
| 4 | Inoculation of agar plates | 10 | |
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Antimicrobial susceptibility testing - EUCAST disk diffusion method, Version 11.0. Växjö, Sweden: 2023.



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PHENOTYPIC AST – DISK DIFFUSION

EUCAST protocol

- How to prepare and store the agar plates
- How to obtain the bacterial inoculum, inoculate the surface of the agar and incubate the plates
 - Incubation at $35 \pm 1^{\circ}$ C during 18 ± 2 hours for Salmonella spp., stacking no more than five agar plates
- How to read the zone diameters
- Lists of situations that require special attention, including the adjustment of medium composition or incubation conditions for certain bacterial species and for certain antimicrobials
 - Complete Appendix dedicated to *Campylobacter* spp.







PHENOTYPIC AST – DISK DIFFUSION

Appendix A Disk diffusion testing of Campylobacter jejuni and coli The following methodology (Table A1) must be adhered to when performing disk diffusion testing of Campylobacter jejuni and coli according to EUCAST. Disk diffusion methodology for Campylobacter jejuni Table A1 and coli Mueller-Hinton agar supplemented with 5% defibrinated horse blood and 20 mg/L β-NAD (MH-F) Medium In order to reduce swarming, the MH-F plates should be dried prior to inoculation (at 20-25°C overnight, or at 35°C, with the lid removed, for 15 min). Inoculum 0.5 McFarland Microaerobic environment 41±1°C 24 h Incubation should result in confluent growth. Some C. coli isolates may not Incubation have sufficient growth after 24 h incubation. These are re-incubated immediately and inhibition zones read after a total of 40-48 h incubation. An incubation temperature of 41±1°C was chosen to create favourable conditions for growth of Campylobacter spp. Read MH-F plates from the front with the lid removed and with reflected light. Zone edges should be read at the point of complete inhibition as judged by Reading the naked eye with the plate held about 30 cm from the eye and at a 45degree angle to the work bench. Quality Campylobacter jejuni ATCC 33560 Control

Antimicrobial susceptibility testing - EUCAST disk diffusion method, Version 11.0. Växjö, Sweden: 2023.



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PHENOTYPIC AST – DISK DIFFUSION

EUCAST protocol

o Quality control of the agar plates and disks

o Control strains

- Same as for the BMD protocol for Salmonella spp.
- Different for Campylobacter spp. → Campylobacter jejuni ATCC 33560

Disk diffusion should not be used for colistin susceptibility testing

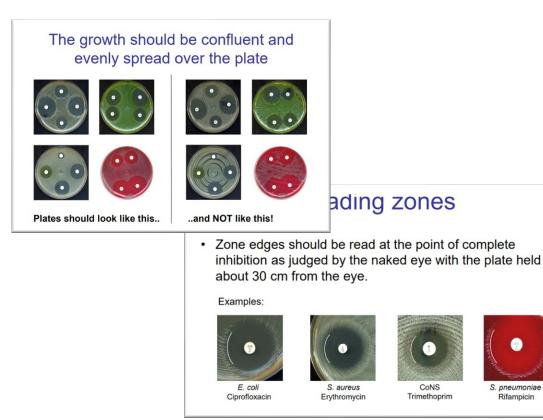




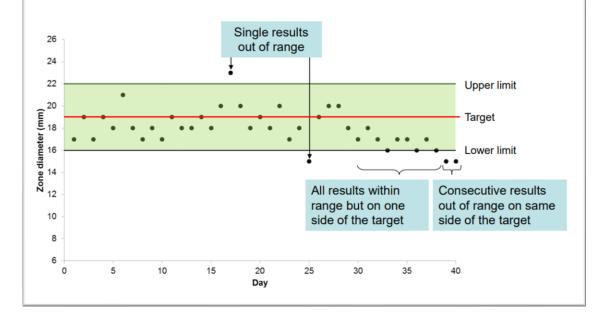


PHENOTYPIC AST – DISK DIFFUSION

EUCAST visual guidelines



Monitoring test performance



EUCAST disk diffusion method for antimicrobial susceptibility testing, Version 11.0. Växjö, Sweden: 2023.







PHENOTYPIC AST – DETECTION OF BETA-LACTAMASES

Phenotypic antimicrobial susceptibility testing

• Broth microdilution

• Disk diffusion



O Detection of β-lactamases

Molecular detection of antimicrobial resistance

- o PCR protocols
- Whole-genome sequencing





PHENOTYPIC AST – DETECTION OF BETA-LACTAMASES

Proposed methods – EUCAST guidelines

- *"EUCAST guidelines for detection of resistance mechanisms and specific resistances ofclinical and/or epidemiological importance. Version 2.0, July 2017"*
- Same as described in "EU protocol for harmonised monitoring of antimicrobial resistance in human Salmonella and Campylobacter isolates"

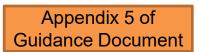


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PHENOTYPIC AST – DETECTION OF BETA-LACTAMASES



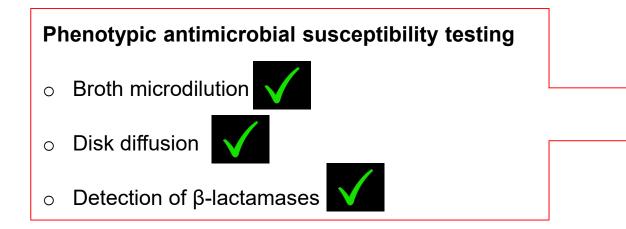
- Production of carbapenemases
 - Tests with meropenem and ertapenem
 - Results are compared with ECOFFs
- \circ Production of extended-spectrum β-lactamases (ESBL)
 - Tests with cefotaxime, ceftazidime or cefepime by themselves and in combination with clavulanic acid
 - Results are compared within themselves (ratio between MIC values / difference between diameters)
- \circ AmpC-mediated β -lactam resistance
 - Tests with cefoxitin, ceftazidime/cefotaxime and cefepime
 - Results are analysed individually (R vs. S)







PHENOTYPIC AST



Concrete examples of documentation for IQC

Molecular detection of antimicrobial resistance

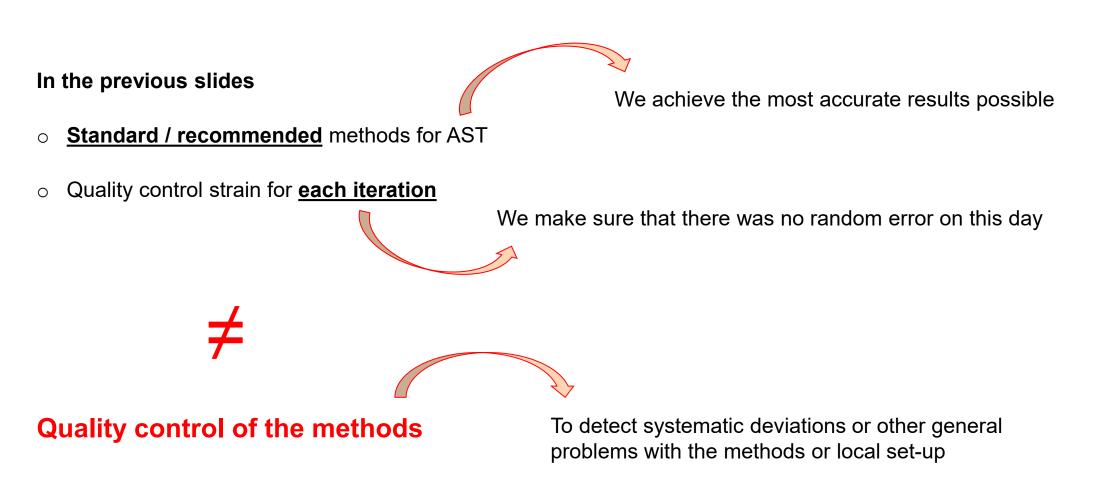
- o PCR protocols
- Whole-genome sequencing







PHENOTYPIC AST – CONCRETE EXAMPLES OF DOCUMENTATION FOR IQC









PHENOTYPIC AST – CONCRETE EXAMPLES OF DOCUMENTATION FOR IQC

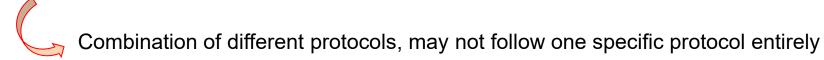
Quality control of the methods

 \circ **Examples** in the Appendices 1 – 4 of the guidance document



Laboratories should not adjust their methods to follow the Appendices, especially if the methods are accredited and/or if they consistenty produce results for control strains within the accepted ranges

Based on <u>DTU</u> SOP





Appendix 1 - Example of method overview documentation for internal quality control

- Ensures all operators follow the same protocol
- Ensures there's no confusion regarding species- or panel- specific details
- Document should be revised as needed (e.g. when purchasing material from different manufacturers)

| Method overview for broth microdilution | | | | | | | | | |
|---|-----------------|----------------------|-----------|--|-------------|--|------------------------------------|--------------------------------------|--------------------|
| Bacteria | Agar | Culture ¹ | MIC panel | Solvent for McFarland suspension | Bouillon | Transfer from McFarland suspension | I noculum to reconstitute wells | Inoculator programme ² | Incubation |
| E. coli | TSA 5% blood | w | EUVSEC3 | dem. water | CAMHB | 10 µl | 50 µl/ well | 1 | 36-37⁰C 18-20 h |
| Salmonella | TSA 5% blood | w | EUVSEC3 | dem. water | CAMHB | 10 µl | 50 µl/ well | 1 | 36-37⁰C 18-20 h |
| Staphylococcus | TSA 5% blood | w | CAMPY | dem. water | CAMHB | 10 µl | 50 µl/ well | 1 | 36-37⁰C 18-20 h |
| Campylobacter ³ | TSA 5% blood | F | CAMPY | CAMHB | CAMH- FB | 100 µl | 50 µl/ well | 25 | 41⁰C 24 h ³ |
| ESBL suspect | TSA 5% blood | w | EUVSEC2 | dem. water | САМНВ | 10 µl | 50 µl/ well | 1 | 36-37⁰C 18-20 h |
| [Other relevant species] | | | | | | | | | |

F: Fresh overnight culture must be used. W: The culture may be refrigerated up to 3 days before use.
 Sensititre autoinoculator equipment number 1234 only.

3) Campylobacter is incubated microaerophilic ($10\% CO_2$, $5\% O_2$, $85\% N_2$) in a CO₂-incubator or anaerobic container. NB: EUCAST recommends $41 \pm 1^{\circ}$ C for 24 hours to achieve better growth and more stable MIC-values, and suggests that isolates with poor growth may be re-incubated and re-read again after a total of 40-48 hours.

Document approved by: Approval date:





FWD AMR· RefLabCap



Appendix 2 - Example of batch of reagents, materials and equipment documentation for internal quality control

- Allows for identification of materialspecific deviations
- Ensures traceability
- Can aid in stock management

| Date/initials | | | |
|----------------------------|--|--|--|
| Dem. water | | | |
| САМНВ | | | |
| CAMH-FB | | | |
| TSA 5% blood | | | |
| EUVSEC3 | | | |
| EUVSEC2 | | | |
| САМРҮ | | | |
| Inoculator 1234 | | | |
| Inoculator 5678 | | | |
| McFarland std. | | | |
| Dosing heads | | | |
| Incubator AB12 | | | |
| Incubator CD34 | | | |
| CO ₂ -incubator | | | |
| [other] | | | |
| [other] | | | |

Batch and equipment documentation for internal guality control of broth microdilution

Remarks:

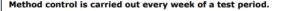




Appendix 3 - Example of method control documentation for internal quality control

- Should be performed regularly to ensure general method conformity
- Should also be performed when new batches of relevant material are received (e.g. new media) – ensures no batch-specific deviations
- Should be revised as needed (e.g. when purchasing material from different manufacturers, and every year when new guidelines are published)

Method control for broth microdilution



Method control is performed for every new batch of panels or media.

Test forms for quality control must be attached. Results sheets for all test isolates must be attached.

Table 1. Reference strains to be used for weekly method control and for control of new batches of panels or media

| Reference strain | | E. coli ATCC 25922 | <i>E. coli</i> NCTC 13846 | <i>C. jejuni</i> ATCC 33560 | S. aureus ATCC 29213 |
|---------------------------|---------|-----------------------|------------------------------|--------------------------------|-------------------------|
| Medi | a | CAMHB | CAMHB | CAMH-FB | CAMHB |
| MIC namel | EUVSEC3 | × | × | | |
| MIC panel (Sensititre™ | EUVSEC2 | × | | | |
|) | САМРҮ | | | x | х |

Table 2. Acceptance intervals (mg/L) for approval of method, panels or media

| | Reference strain | | | | | | | |
|----------------|-----------------------|-----------------------|-------------------------|-------------------------|--|--|--|--|
| Antimicrobials | E. coli ATCC 25922 | E. coli NCTC 13846 | C. jejuni ATCC 33560 | S. aureus ATCC 29213 | | | | |
| Amikacin | 0.5-4 | | | 1-4 | | | | |
| Ampicillin | 2-8 | | | | | | | |
| [] | [] | [] | [] | [] | | | | |
| Ciprofloxacin | 0.004-0.016 | | | 0.125-0.5 | | | | |
| Clindamycin | | | | 0.06-0.25 | | | | |
| Colistin | 0.25-2 | 2-8 | | | | | | |
| [] | [] | [] | [] | [] | | | | |

| Purpose: [] weekly control | [] panel batch control |] media batch control | [] |
|----------------------------|------------------------|---|----|
| Panel code: | | | |
| Panel batch: | | | |
| Panel expiration date: | | | |
| Broth code: | | | |
| Broth batch: | | | |
| Broth expiration date: | | | |
| Desferring discus | | | |
| Performed by: | _ | | |
| Date: | | | |
| Read by: | | | |
| Date: | | | |
| | | | |





Appendix 4 - Example of documentation for quality control of each AST <u>iteration</u> ("test form")

- There should be a "test form" for each combination of control strain + panel
- Allows for quick evaluation of conformity with accepted ranges
- Allows for long-term evaluation of trends in deviations
- Avoids errors because accepted range is coloured
- Should be revised for new combinations of QC strains+panels, and every year when new guidelines are published

"Test form" for quality control for broth microdilution

Quality control is carried out at least once a day when testing is performed.

Control strain: *Escherichia coli* ATCC 25922 Panel: EUVSEC3 Broth medium: CAMHB Volume per well: 50 µl Accepted ranges: Green (EUCAST QC tables v13.0, valid from 01/01/2023)

| 12 | 11 | 10 | 9 | 8 | 7 | 6 | 5 | 4 | 3 | 2 | 1 | |
|-----|------|------|------|------|------|------|------|------|------|------|-------|---|
| SMX | TMP | TET | NAL | COL | FOT | TAZ | TGC | GEN | AMI | AZI | AMP | Α |
| 512 | 16 | 32 | 64 | 16 | 4 | 8 | 8 | 16 | 128 | 64 | 32 | |
| SMX | TMP | TET | NAL | COL | FOT | TAZ | TGC | GEN | AMI | AZI | AMP | в |
| 256 | 8 | 16 | 32 | 8 | 2 | 4 | 4 | 8 | 64 | 32 | 16 | |
| SMX | TMP | TET | NAL | COL | FOT | TAZ | TGC | GEN | AMI | AZI | AMP | С |
| 128 | 4 | 8 | 16 | 4 | 1 | 2 | 2 | 4 | 32 | 16 | 8 | |
| SMX | TMP | TET | NAL | COL | FOT | TAZ | TGC | GEN | AMI | AZI | AMP | D |
| 64 | 2 | 4 | 8 | 2 | 0.5 | 1 | 1 | 2 | 16 | 8 | 4 | |
| SMX | TMP | TET | NAL | COL | FOT | TAZ | TGC | GEN | AMI | AZI | AMP | Е |
| 32 | 1 | 2 | 4 | 1 | 0.25 | 0.5 | 0.5 | 1 | 8 | 4 | 2 | |
| SMX | TMP | CHL | CHL | CHL | CHL | TAZ | TGC | GEN | AMI | AZI | AMP | F |
| 16 | 0.5 | 64 | 32 | 16 | 8 | 0.25 | 0.25 | 0.5 | 4 | 2 | 1 | |
| SMX | TMP | MERO | G |
| 8 | 0.25 | 16 | 8 | 4 | 2 | 1 | 0.5 | 0.25 | 0.12 | 0.06 | 0.03 | |
| POS | POS | CIP | н |
| CON | CON | 8 | 4 | 2 | 1 | 0.5 | 0.25 | 0.12 | 0.06 | 0.03 | 0.015 | |
| ; | POS | CIP | н |

| Code AMI AMP AZI FOT TAZ CHL CIP COL GEN MERO NAI | Antimicrobial agent (15) AMIKACIN AMICILLIN AZITHROMYCIN CEFOTAXIME CEFTAZIDIME CHLORAMPHENICOL CIPROFLOXACIN COLISTIN GENTAMICIN MEROPENEM NAI IDIXIC ACID | Test range (mg/L) 4-128 1-32 2-64 0.25-4 0.25-8 8-64 0.015-8 1-16 0.5-16 0.03-16 4-64 | | | | | |
|--|--|---|--|--|--|--|--|
| SMX | SULFAMETHOXAZOLE | 8-512 | | | | | |
| TET | TETRACYCLINE | 2-32 | | | | | |
| TGC | TIGECYCLINE | 0.25-8 | | | | | |
| TMP | TRIMETHOPRIM | 0.25-16 | | | | | |
| POS | POSITIVE CONTROL | 2x | | | | | |
| Performed by: Date: | | | | | | | |
| Read by: Date: | | | | | | | |
| Remarks: | | | | | | | |

A similar layout <u>without</u> colour and <u>without</u> having the headers pre-completed can be used to record the actual test results for each isolate

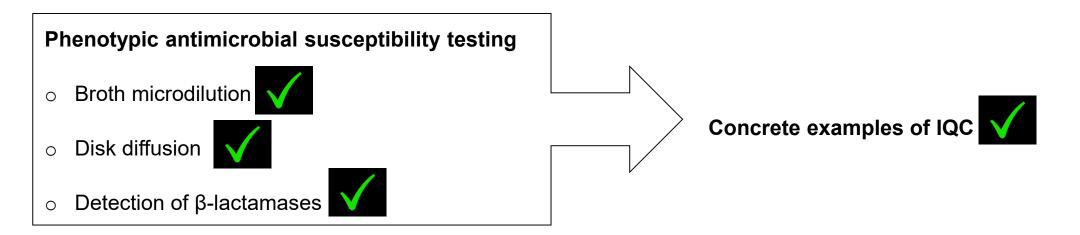








PHENOTYPIC AST – PCR PROTOCOLS



Molecular detection of antimicrobial resistance

- PCR protocols
- Whole-genome sequencing





MOLECULAR DETECTION – PCR PROTOCOLS



Appendix 6 of Guidance Document

Molecular detection of resistance in Salmonella spp.

o Databases: Beta-Lactamase Database (BLDB), EURL-AR list of *mcr*-genes, ResFinder, other

- Acquired AMR genes \rightarrow PCR protocols
 - EuSCAPE multiplex PCR protocol (*bla*_{KPC}, *bla*_{VIM}, *bla*_{OXA-48} and *bla*_{NDM})
 - Dallene et al. set of 6+1 multiplex/simplex PCR (*bla*_{TEM}, *bla*_{SHV}, *bla*_{OXA}, *bla*_{CTX-M}, *bla*_{VIM}, *bla*_{IMP}, *bla*_{KPC}, *bla*_{VEB}, *bla*_{GES}, *bla*_{PER} and plasmid-mediated AmpC β-lactamases)
 - EURL-AR multiplex PCR protocol (*mcr-1* to *mcr-5*)
 - Borowiak et al. multiplex PCR protocol (*mcr-6* to *mcr-9*)
- Chromosomal point mutations → sequencing







MOLECULAR DETECTION – PCR PROTOCOLS

Molecular detection of resistance in Campylobacter spp.

Databases: ResFinder, literature, other(?)

- Acquired AMR genes \rightarrow PCR protocols
 - Eryıldız et al. multiplex PCR protocol (*tet(O*) and *erm(B*))

○ Chromosomal point mutations \rightarrow sequencing







MOLECULAR DETECTION – PCR PROTOCOLS

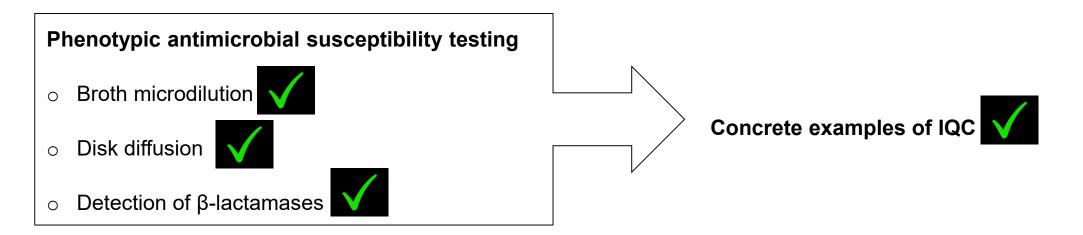
- Quality control
 - Use all positive control strains described in the chosen PCR protocol
 - Always include a negative control
 - Do not combine different PCR protocols into a larger multiplex
 - Do not use terms "susceptible" or "resistant" → report results as presence or absence of the genes included in the protocols
 - Create method overview documentation and record the batch of reagents, materials and equipment







PHENOTYPIC AST – WHOLE-GENOME SEQUENCING



Molecular detection of antimicrobial resistance

• PCR protocols



• Whole-genome sequencing







MOLECULAR DETECTION – WHOLE-GENOME SEQUENCING

Molecular detection resistance through WGS

• Databases: Same as before, other bioinformatics tools and databases such as CARD-RGI and AMRFinderPlus

- Techniques described in FWD AMR-RLC WGS protocol
 - Harmonisation of QC is difficult due to diversity of options
 - Main QC parameters and respective thresholds proposed in the protocol





STATENS SERUM IN STITUT

INTERNAL QUALITY CONTROL – FOR AST

MOLECULAR DETECTION - WHOLE-GENOME SEQUENCING

- Quality control
 - Create method overview documentation and record the batch of reagents, materials and equipment
 - Record the version and/or date of the bioinformatics tools and databases that are used for analysis of raw sequence data
 - Store the raw sequence data permanently
 - Apply well-defined QC thresholds for raw data and for assemblies **

** For raw sequence data:

• Average read length: Should correspond to that expected from the sequencing platform and kit (e.g. Illumina NextSeq read length is approximately 150 base-pairs (bp));

buld be contaminated.

- Number of reads: Should be as high as possible and at least enough to obtain the desired depth of coverage;
- Depth of coverage: Should as a minimum be 30X, and
 Species identification: If more than 5% of the reads ma

Appendix 7 of

** For assembled data:

Size of assembled genome: Should be within the range for the targeted organism (4.4-5.8 million bp for Salmonella and 1.5-1.9 million bp for Campylobacter);

Guidance Document

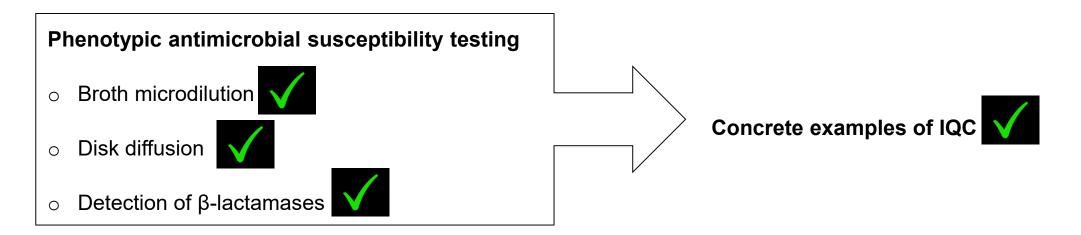
- N50: Should be higher than 30,000 bp;
- Total number of contigs: Should be less than 500 (Campylobacter will typically be assembled into less than 100 contigs and Salmonella to less than 300 contigs).







PHENOTYPIC AST – WHOLE-GENOME SEQUENCING



Molecular detection of antimicrobial resistance

• PCR protocols



• Whole-genome sequencing









Questions and discussion







Thank you on behalf of the FWD AMR-RefLabCap team

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