



HaDEA Service Contract 20197409

Provision of EU networking and support for public health reference laboratory functions for antimicrobial resistance in *Salmonella* species and *Campylobacter* species in human samples







Webinar

Guidance document on internal quality control schemes for reference antimicrobial susceptibility testing and detection of genetic determinants of antimicrobial resistance for *Salmonella* and *Campylobacter* isolates from human samples

Wednesday, 22 March 2023, 14:00-15:00 CET

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Virtual Housekeeping



Please **turn off your cameras and microphones** unless you're speaking – this will help with bandwidth and maximise audibility.



Do frequently **use the chat function** to share your views, comments and challenges. Keep the chat constructive, respectful and on topic!



If you wish to make a comment for e.g. the discussion, please use the 'Raise hand' function.







Meeting agenda

- 1. Presentation of the guidance document
- Background and aim
- IQC in general
- IQC for AST

2. Discussion





BACKGROUND AND AIM











BACKGROUND AND AIM



- Collect the most recently available information from different regulatory agencies and other sources
- Aid the NRLs in optimizing their current methods and implementing molecular methods
- Provide concrete examples of techniques for Internal Quality Control
- Allow the NRLs to easily provide guidance or training to local national laboratories
- Describe the standardized / recommended methods for antimicrobial susceptibility testing

Reliable and accurate results for diagnostics and surveillance purposes of Salmonella and Campylobacter

Data that are comparable within Europe for surveillance purposes





BACKGROUND AND AIM



TABLE OF CONTENTS

1.	INTRODUCTION AND SCOPE	4
2.	INTERNAL QUALITY CONTROL STRATEGIES	4
3.	INTERNAL QUALITY CONTROL WHEN PERFORMING PHENOTYPIC ANTIMICROBIAL SUSCEPTIBILITY TESTING	6
	3.1. Background	
	3.3. Disk diffusion	
	3.4. Quality control of phenotypic antimicrobial susceptibility testing3.5. Phenotypic detection of β-lactamase-producing bacteria	
4.	INTERNAL QUALITY CONTROL WHEN PERFORMING MOLECULAR DETECTION OF ANTIMICROBIAL RESISTANCE DETERMINANTS	. 11
5.	REFERENCES	. 12
6.	APPENDICES	. 17
ſ	6.1. Appendix 1	
	6.2. Appendix 2	. 18
	6.3. Appendix 3	. 19
Į	6.4. Appendix 4	
	6.5. Appendix 5	. 21
=	6.6. Appendix 6	. 22
	6.7. Appendix 7	





Concrete examples of IQC documentation

Methods descriptions





ISO standards

- o ISO 15189
- ISO/IEC 17025



- Promote internal quality and competence
- Facilitate internal and external comparibility of results



EQA and accreditation

- External quality assessment exercises
- Strategy for accreditation



Increase confidence regarding accuracy of results









ISO STANDARDS – ISO 15189

ISO standards

- o ISO 15189
- o ISO/IEC 17025

EQA and accreditation

- External quality assessment exercises
- Strategy for accreditation







ISO STANDARDS – ISO 15189

ISO 15189:2022

"Medical laboratories - Requirements for quality and competence"

Deep re-organization of the document compared with :2012 version

:2012	:2022
1. 2. 3. 4. 5.	1. 2. 3. 4. 5. 6. 7.
	0.









ISO STANDARDS - ISO 15189







ISO STANDARDS - ISO 15189







ISO STANDARDS – ISO 15189

Example: 6) Resource requirements; 6.5) Equipment calibration and metrological traceability







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ISO STANDARDS – ISO 15189

Example: 7) Process requirements; 7.3) Examination processes









ISO STANDARDS – ISO 15189

Example: 8) Management system requirements; 8.4) Control of records









ISO STANDARDS - ISO/IEC 17025

ISO standards



ISO/IEC 17025

EQA and accreditation

- External quality assessment exercises
- Strategy for accreditation







ISO STANDARDS – ISO/IEC 17025

ISO/IEC 17025:2017

"General requirements for the competence of testing and calibration laboratories"









ISO STANDARDS – ISO/IEC 17025







ISO STANDARDS – ISO/IEC 17025

Now more similar to ISO 15189:2022









EXTERNAL QUALITY ASSESSMENT EXERCISES

ISO standards



o ISO/IEC 17025

EQA and accreditation

- External quality assessment exercises
- Strategy for accreditation









EXTERNAL QUALITY ASSESSMENT EXERCISES

Examples:

EARS-Net EQA / ECDC's EQA on AST / UK NEQAS / ESfEQA / Labquality / Oneworld Accuracy

Recommended within the ISO standards







STRATEGY FOR ACCREDITATION

ISO standards

o ISO 15189

o ISO/IEC 17025

EQA and accreditation

External quality assessment exercises



Strategy for accreditation









STRATEGY FOR ACCREDITATION

International Laboratory Accreditation Cooperation (ILAC)

L Specifically designated body - national institution

L Evaluation and accreditation of reference/local laboratories





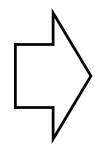




ISO standards

o ISO 15189

○ ISO/IEC 17025



Internal quality control for AST

EQA and accreditation

External quality assessment exercises



Strategy for accreditation









European guidance on AST methods:

European Committee on Antimicrobial Susceptibility Testing (EUCAST)

Recommendations:

- Broth microdilution or disk diffusion for AST
- Other methods (agar dilution / gradient strips) are not recommended due to lack of harmonisation and high variability
- Regularly confirming warnings and new breakpoint tables







Phenotypic antimicrobial susceptibility testing

- Broth microdilution
- Disk diffusion
- Detection of β-lactamases

Molecular detection of antimicrobial resistance

- o PCR protocols
- Whole-genome sequencing







PHENOTYPIC AST – BROTH MICRODILUTION

Phenotypic antimicrobial susceptibility testing

- Broth microdilution
- Disk diffusion
- Detection of β-lactamases

Molecular detection of antimicrobial resistance

- o PCR protocols
- Whole-genome sequencing









PHENOTYPIC AST – BROTH MICRODILUTION

Standard protocol – ISO 20776-1:2019

 "Susceptibility testing of infectious agents and evaluation of performance of antimicrobial susceptibility test devices -Part 1: Broth micro-dilution reference method for testing the in vitro activity of antimicrobial agents against rapidly growing aerobic bacteria involved in infectious diseases"

EUCAST documents

- Clinical breakpoint tables
- Warnings page
- Visual guides (e.g. how to determine MIC endpoints)







PHENOTYPIC AST – BROTH MICRODILUTION

ISO 20776-1:2019









PHENOTYPIC AST – BROTH MICRODILUTION

ISO 20776-1:2019

- o How to prepare stock and working solutions of antimicrobial agents, the broth medium and the microdilution trays
- Two methods for obtaining the bacterial inoculum: the broth culture method and the direct colony suspension method
 - Final concentration of 5 x 10⁵ CFU/ml
- o How to inoculate, incubate and read the minimum inhibitory concentrations (MIC) on the microdilution trays
- Lists of situations that require special attention, including the adjustment of medium composition or incubation conditions for certain bacterial species and for certain antimicrobials









PHENOTYPIC AST – BROTH MICRODILUTION

Examples of special situations (ISO 20776-1 + EUCAST)

- For Salmonella spp.
 - Do not add surfactants to the medium when testing colistin
 - Adjust the zinc concentration of the broth medium for testing of carbapenems

- For Campylobacter spp.
 - Use MH-F broth (MH broth supplemented with lysed horse blood and β-NAD)
 - Incubate under the special conditions of 41± 1°C during 24 hours in microaerobic environment, and extension of incubation time up to 40-48 hours might be necessary if growth is insufficient







PHENOTYPIC AST – BROTH MICRODILUTION

- Use of control strains
 - List from the Clinical Laboratory Standards Institute (CLSI) (available on the document CLSI M100 "Performance Standards for Antimicrobial Susceptibility Testing")



 List from EUCAST (available on the document "Routine and extended internal quality control for MIC determination and disk diffusion as recommended by EUCAST")



Salmonella spp.

Always: Escherichia coli ATCC 25922

Colistin: mcr-1-positive E. coli NCTC 13846

β-lactams+ β-lactamase inhibitors: *E. coli* ATCC 35218, others

Campylobacter spp.

Always: Staphylococcus aureus ATCC 29213

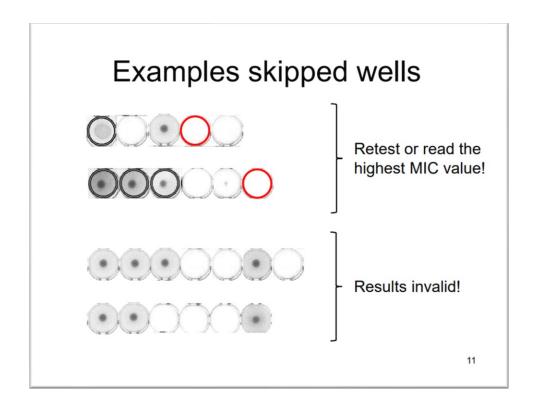






PHENOTYPIC AST – BROTH MICRODILUTION

EUCAST visual guidelines



Trimethoprim and trimethoprim-sulfamethoxazole Read the MIC at the lowest concentration that inhibits ≥80 % of growth as compared to the growth control. Growth control 16









PHENOTYPIC AST - DISK DIFFUSION

Phenotypic antimicrobial susceptibility testing

Broth microdilution

- Disk diffusion
- Detection of β-lactamases

Molecular detection of antimicrobial resistance

- o PCR protocols
- Whole-genome sequencing







PHENOTYPIC AST - DISK DIFFUSION

Standard protocol – EUCAST protocol

o "Antimicrobial susceptibility testing - EUCAST disk diffusion method. Version 11.0, January 2023"

EUCAST documents

- Clinical breakpoint tables
- Warnings page
- Visual guides (e.g. how to confirm adequate growth and determine zone diameters)







PHENOTYPIC AST – DISK DIFFUSION

EUCAST protocol

Contents		Page	
	Changes from previous version		
	Abbreviations and Terminology		
1	Introduction	5	
2	Preparation and storage of media	6	
3	Preparation of inoculum	8	
4	Inoculation of agar plates	10	
5	Application of antimicrobial disks	11	
6	Incubation of plates	12	
7	Examination of plates after incubation	14	
8	Measurement of zones and interpretation of susceptibility	15	
9	Quality control	17	
	Appendix A	21	

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PHENOTYPIC AST – DISK DIFFUSION

EUCAST protocol

- How to prepare and store the agar plates
- How to obtain the bacterial inoculum, inoculate the surface of the agar and incubate the plates
 - Incubation at $35 \pm 1^{\circ}$ C during 18 ± 2 hours for *Salmonella* spp., stacking no more than five agar plates
- How to read the zone diameters
- Lists of situations that require special attention, including the adjustment of medium composition or incubation conditions for certain bacterial species and for certain antimicrobials
 - Complete Appendix dedicated to Campylobacter spp.







PHENOTYPIC AST – DISK DIFFUSION

Appendix A

Disk diffusion testing of Campylobacter jejuni and coli

The following methodology (Table A1) must be adhered to when performing disk diffusion testing of Campylobacter jejuni and coli according to EUCAST.

Table A1	Disk diffusion methodology for <i>Campylobacter jejuni</i> and <i>coli</i>
Medium	Mueller-Hinton agar supplemented with 5% defibrinated horse blood and 20 mg/L β-NAD (MH-F) In order to reduce swarming, the MH-F plates should be dried prior to inoculation (at 20-25°C overnight, or at 35°C, with the lid removed, for 15 min).
Inoculum	0.5 McFarland
Incubation	Microaerobic environment 41±1°C 24 h Incubation should result in confluent growth. Some <i>C. coli</i> isolates may not have sufficient growth after 24 h incubation. These are re-incubated immediately and inhibition zones read after a total of 40-48 h incubation. An incubation temperature of 41±1°C was chosen to create favourable conditions for growth of <i>Campylobacter</i> spp.
Reading	Read MH-F plates from the front with the lid removed and with reflected light. Zone edges should be read at the point of complete inhibition as judged by the naked eye with the plate held about 30 cm from the eye and at a 45-degree angle to the work bench.
Quality Control	Campylobacter jejuni ATCC 33560

FWD AMR.



Antimicrobial susceptibility testing - EUCAST disk diffusion method, Version 11.0. Växjö, Sweden: 2023.





PHENOTYPIC AST - DISK DIFFUSION

EUCAST protocol

Quality control of the agar plates and disks

- Control strains
 - Same as for the BMD protocol for Salmonella spp.
 - Different for Campylobacter spp. → Campylobacter jejuni ATCC 33560

Disk diffusion should not be used for colistin susceptibility testing

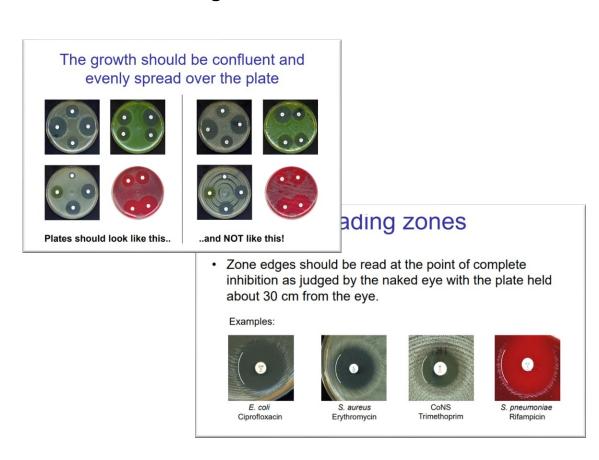


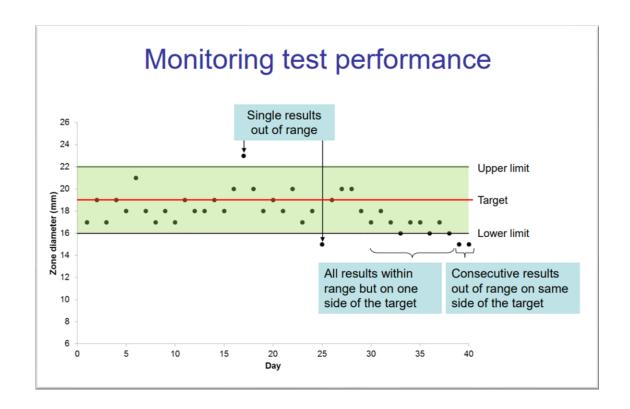




PHENOTYPIC AST – DISK DIFFUSION

EUCAST visual guidelines







EUCAST disk diffusion method for antimicrobial susceptibility testing, Version 11.0. Växjö, Sweden: 2023.





PHENOTYPIC AST – DETECTION OF BETA-LACTAMASES

Phenotypic antimicrobial susceptibility testing

- Broth microdilution
- Disk diffusion
- Detection of β-lactamases

Molecular detection of antimicrobial resistance

- PCR protocols
- Whole-genome sequencing









PHENOTYPIC AST – DETECTION OF BETA-LACTAMASES

Proposed methods – EUCAST guidelines

- "EUCAST guidelines for detection of resistance mechanisms and specific resistances ofclinical and/or epidemiological importance. Version 2.0, July 2017"
- Same as described in "EU protocol for harmonised monitoring of antimicrobial resistance in human Salmonella and Campylobacter isolates"







Appendix 5 of Guidance Document

PHENOTYPIC AST – DETECTION OF BETA-LACTAMASES

- Production of carbapenemases
 - Tests with meropenem and ertapenem
 - Results are compared with ECOFFs
- Production of extended-spectrum β-lactamases (ESBL)
 - Tests with cefotaxime, ceftazidime or cefepime by themselves and in combination with clavulanic acid
 - Results are compared within themselves (ratio between MIC values / difference between diameters)
- AmpC-mediated β-lactam resistance
 - Tests with cefoxitin, ceftazidime/cefotaxime and cefepime
 - Results are analysed individually (R vs. S)









PHENOTYPIC AST

Phenotypic antimicrobial susceptibility testing

Broth microdilution

Disk diffusion



Concrete examples of documentation for IQC

Molecular detection of antimicrobial resistance

- PCR protocols
- Whole-genome sequencing







PHENOTYPIC AST – CONCRETE EXAMPLES OF DOCUMENTATION FOR IQC

In the previous slides

We achieve the most accurate results possible

- Standard / recommended methods for AST
- Quality control strain for <u>each iteration</u>

We make sure that there was no random error on this day





Quality control of the methods

To detect systematic deviations or other general problems with the methods or local set-up









PHENOTYPIC AST – CONCRETE EXAMPLES OF DOCUMENTATION FOR IQC

Quality control of the methods

Examples in the Appendices 1 - 4 of the guidance document



Laboratories should not adjust their methods to follow the Appendices, especially if the methods are accredited and/or if they consistenty produce results for control strains within the accepted ranges

Based on **DTU** SOP



Combination of different protocols, may not follow one specific protocol entirely



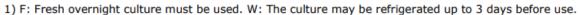


Appendix 1 - Example of method overview documentation for internal quality control

- Ensures all operators follow the same protocol
- Ensures there's no confusion regarding species- or panel- specific details
- Document should be revised as needed (e.g. when purchasing material from different manufacturers)

Method overview for broth microdilution

Bacteria	Agar	Culture 1	MIC panel	Solvent for McFarland suspension	Bouillon	Transfer from McFarland suspension	Inoculum to reconstitute wells	Inoculator programme ²	Incubation
E. coli	TSA 5% blood	w	EUVSEC3	dem. water	САМНВ	10 µl	50 µl/ well	1	36-37°C 18-20 h
Salmonella	TSA 5% blood	w	EUVSEC3	dem. water	САМНВ	10 µl	50 µl/ well	1	36-37°C 18-20 h
Staphylococcus	TSA 5% blood	w	CAMPY	dem. water	САМНВ	10 µl	50 µl/ well	1	36-37°C 18-20 h
Campylobacter ³	TSA 5% blood	F	CAMPY	САМНВ	CAMH- FB	100 μl	50 µl/ well	25	41°C 24 h ³
ESBL suspect	TSA 5% blood	w	EUVSEC2	dem. water	САМНВ	10 μΙ	50 µl/ well	1	36-37°C 18-20 h
[Other relevant species]									



²⁾ Sensititre autoinoculator equipment number 1234 only.

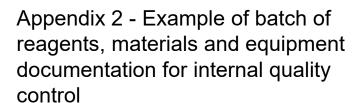
Document approved by: Approval date:





³⁾ Campylobacter is incubated microaerophilic (10% CO $_2$, 5% O $_2$, 85% N $_2$) in a CO $_2$ -incubator or anaerobic container. NB: EUCAST recommends $41 \pm 1\%$ for 24 hours to achieve better growth and more stable MIC-values, and suggests that isolates with poor growth may be re-incubated and re-read again after a total of 40-48 hours.





- Allows for identification of materialspecific deviations
- Ensures traceability
- Can aid in stock management



Batch and equipment	documentation for	or internal quality	control of br	oth microdilution

Table 1. Batch of reagents, materials and equipment

Batch and equipment control is carried out for every test iteration.

Date/initials			
Dem. water			
САМНВ			
CAMH-FB			
TSA 5% blood			
EUVSEC3			
EUVSEC2			
CAMPY			
Inoculator 1234			
Inoculator 5678			
McFarland std.			
Dosing heads			
Incubator AB12			
Incubator CD34			
CO ₂ -incubator			
[other]			
[other]			

Remarks:





Appendix 3 - Example of method control documentation for internal quality control

- Should be performed regularly to ensure general method conformity
- Should also be performed when new batches of relevant material are received (e.g. new media) – ensures no batch-specific deviations
- Should be revised as needed (e.g. when purchasing material from different manufacturers, and every year when new guidelines are published)

Method control for broth microdilution

Method control is carried out every week of a test period.

Method control is performed for every new batch of panels or media.

Test forms for quality control must be attached. Results sheets for all test isolates must be attached.

Table 1. Reference strains to be used for weekly method control and for control of new batches of panels or media

Reference	strain	E. coli ATCC 25922	E. coli NCTC 13846	C. jejuni ATCC 33560	S. aureus ATCC 29213
Media		САМНВ	САМНВ	CAMH-FB	САМНВ
MIChanal	EUVSEC3	×	×		
MIC panel (Sensititre™	EUVSEC2	×			
)	CAMPY			X	Х

Table 2. Acceptance intervals (mg/L) for approval of method, panels or media

		Reference strain							
Antimicrobials	E. coli ATCC 25922	E. coli NCTC 13846	C. jejuni ATCC 33560	S. aureus ATCC 29213					
Amikacin	0.5-4			1-4					
Ampicillin	2-8								
[]	[]	[]	[]	[]					
Ciprofloxacin	0.004-0.016			0.125-0.5					
Clindamycin				0.06-0.25					
Colistin	0.25-2	2-8							
[]	[]	[]	[]	[]					

Purpose: [] weekly control	[] panel batch control	[] media batch control	[]
Panel code:			
Panel batch:			
Panel expiration date:			
Broth code:			
Broth batch:			
Broth expiration date:			
Performed by: Date:	_		
Read by:			
Date:			
Remarks:			







Appendix 4 - Example of documentation for quality control of each AST <u>iteration</u> ("test form")

- There should be a "test form" for each combination of control strain + panel
- Allows for quick evaluation of conformity with accepted ranges
- Allows for long-term evaluation of trends in deviations
- Avoids errors because accepted range is coloured
- Should be revised for new combinations of QC strains+panels, and every year when new guidelines are published

"Test form" for quality control for broth microdilution

Quality control is carried out at least once a day when testing is performed.

Control strain: Escherichia coli ATCC 25922

Panel: EUVSEC3 Broth medium: CAMHB Volume per well: 50 µl

Remarks:

Accepted ranges: Green (EUCAST QC tables v13.0, valid from 01/01/2023)

	1	2	3	4	5	6	7	8	9	10	11	12
Α	AMP	AZI	AMI	GEN	TGC	TAZ	FOT	COL	NAL	TET	TMP	SMX
	32	64	128	16	8	8	4	16	64	32	16	512
В	AMP	AZI	AMI	GEN	TGC	TAZ	FOT	COL	NAL	TET	TMP	SMX
	16	32	64	8	4	4	2	8	32	16	8	256
С	AMP	AZI	AMI	GEN	TGC	TAZ	FOT	COL	NAL	TET	TMP	SMX
	8	16	32	4	2	2	1	4	16	8	4	128
D	AMP	AZI	AMI	GEN	TGC	TAZ	FOT	COL	NAL	TET	TMP	SMX
	4	8	16	2	1	1	0.5	2	8	4	2	64
E	AMP 2	AZI 4	AMI 8	GEN 1	TGC 0.5	TAZ 0.5	FOT 0.25	COL 1	NAL 4	TET 2	TMP 1	SMX 32
F	AMP	AZI	AMI	GEN	TGC	TAZ	CHL	CHL	CHL	CHL	TMP	SMX
	1	2	4	0.5	0.25	0.25	8	16	32	64	0.5	16
G	MERO	MERO	MERO	MERO	MERO	MERO	MERO	MERO	MERO	MERO	TMP	SMX
	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	0.25	8
н	CIP	CIP	CIP	CIP	CIP	CIP	CIP	CIP	CIP	CIP	POS	POS
	0.015	0.03	0.06	0.12	0.25	0.5	1	2	4	8	CON	CON

Code	Antimicrobial agent (15)	Test range (mg/L)					
AMI	AMIKACIN	4-128					
AMP	AMPICILLIN	1-32					
AZI	AZITHROMYCIN	2-64					
FOT	CEFOTAXIME	0.25-4					
TAZ	CEFTAZIDIME	0.25-8					
CHL	CHLORAMPHENICOL	8-64					
CIP	CIPROFLOXACIN	0.015-8					
COL	COLISTIN	1-16					
GEN	GENTAMICIN	0.5-16					
MERO	MEROPENEM	0.03-16					
NAL	NALIDIXIC ACID	4-64					
SMX	SULFAMETHOXAZOLE	8-512					
TET	TETRACYCLINE	2-32					
TGC	TIGECYCLINE	0.25-8					
TMP	TRIMETHOPRIM	0.25-16					
POS	POSITIVE CONTROL	2x					
Performed by:							
Date:							
Read by: Date:							

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A similar layout without colour and without having the headers pre-completed can be used to record the actual test results for each isolate

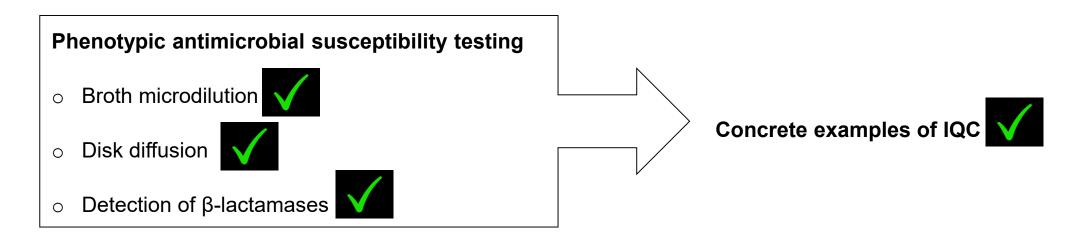








PHENOTYPIC AST – PCR PROTOCOLS



Molecular detection of antimicrobial resistance

- PCR protocols
- Whole-genome sequencing







MOLECULAR DETECTION – PCR PROTOCOLS

Appendix 6 of Guidance Document

Molecular detection of resistance in Salmonella spp.

o Databases: Beta-Lactamase Database (BLDB), EURL-AR list of *mcr*-genes, ResFinder, other

- Acquired AMR genes → PCR protocols
 - EuSCAPE multiplex PCR protocol (bla_{KPC}, bla_{VIM}, bla_{OXA-48} and bla_{NDM})
 - Dallene et al. set of 6+1 multiplex/simplex PCR (bla_{TEM}, bla_{SHV}, bla_{OXA}, bla_{CTX-M}, bla_{VIM}, bla_{IMP}, bla_{KPC}, bla_{VEB}, bla_{GES}, bla_{PER} and plasmid-mediated AmpC β-lactamases)
 - EURL-AR multiplex PCR protocol (*mcr-1* to *mcr-5*)
 - Borowiak et al. multiplex PCR protocol (mcr-6 to mcr-9)
- Chromosomal point mutations → sequencing







MOLECULAR DETECTION – PCR PROTOCOLS

Molecular detection of resistance in Campylobacter spp.

Databases: ResFinder, literature, other(?)

- Acquired AMR genes → PCR protocols
 - Eryildiz et al. multiplex PCR protocol (tet(O) and erm(B))

○ Chromosomal point mutations → sequencing







MOLECULAR DETECTION – PCR PROTOCOLS

- Quality control
 - Use all positive control strains described in the chosen PCR protocol
 - Always include a negative control
 - Do not combine different PCR protocols into a larger multiplex
 - Do not use terms "susceptible" or "resistant" → report results as presence or absence of the genes included in the protocols
 - Create method overview documentation and record the batch of reagents, materials and equipment







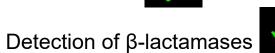


PHENOTYPIC AST – WHOLE-GENOME SEQUENCING

Phenotypic antimicrobial susceptibility testing

Broth microdilution

Disk diffusion



Concrete examples of IQC



Molecular detection of antimicrobial resistance

PCR protocols

Whole-genome sequencing







MOLECULAR DETECTION — WHOLE-GENOME SEQUENCING

Molecular detection resistance through WGS

o Databases: Same as before, other bioinformatics tools and databases such as CARD-RGI and AMRFinderPlus

- Techniques described in FWD AMR-RLC WGS protocol
 - Harmonisation of QC is difficult due to diversity of options
 - Main QC parameters and respective thresholds proposed in the protocol







MOLECULAR DETECTION – WHOLE-GENOME SEQUENCING

- Quality control
 - Create method overview documentation and record the batch of reagents, materials and equipment
 - Record the version and/or date of the bioinformatics tools and databases that are used for analysis of raw sequence data
 - Store the raw sequence data permanently
 - Apply well-defined QC thresholds for raw data and for assemblies **

Appendix 7 of
Guidance Document

** For assembled data:

- Size of assembled genome: Should be within the range for the targeted organism (4.4-5.8 million bp for Salmonella and 1.5-1.9 million bp for Campylobacter);
- N50: Should be higher than 30,000 bp;
- Total number of contigs: Should be less than 500 (Campylobacter will typically be assembled into less than 100 contigs and Salmonella to less than 300 contigs).



^{**} For raw sequence data:

Average read length: Should correspond to that expected from the sequencing platform and kit (e.g. Illumina NextSeq read length is approximately 150 base-pairs (bp));

Number of reads: Should be as high as possible and at least enough to obtain the desired depth of coverage;

[■] Depth of coverage: Should as a minimum be 30X, and

Species identification: If more than 5% of the reads ma





PHENOTYPIC AST – WHOLE-GENOME SEQUENCING

Phenotypic antimicrobial susceptibility testing

Broth microdilution

Disk diffusion



Detection of β-lactamases



Concrete examples of IQC



Molecular detection of antimicrobial resistance

p PCR protocols

Whole-genome sequencing









Questions and discussion







Thank you on behalf of the FWD AMR-RefLabCap team

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